A SHORT ROUTE TO THE PHTHALIDEISOQUINOLINES AND THE 13-HYDROXYLATED PROTOBERBERINES¹

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Abstract—CrO₃ oxidation of 2'-hydroxymethylpapaverine (2) yields the aromatic phthalideisoquinoline 3. Catalytic reduction of 3 gives the *erythro*-norphthalideisoquinoline 7 and the *threo* analog 9. Respective N-methylation furnishes the *erythro*-phthalideisoquinoline 8, and the *threo* isomer 10. The nor compounds 7 and 9 can be converted in hydroxylic base to the lactam alcohols 11 and 12, respectively; but 12 tends to dehydrate to the oxyprotoberberine 15. LAH reduction of 11 and 12 affords in turn the 13-hydroxyprotoberberines 13 and 14. Three factors affect the conformation of the phthalideisoquinolines, namely the relative stereochemistry at C-1 and C-9, substitution on nitrogen, and substitution at C-8.

The classical phthalideisoquinoline alkaloids possess a tetracyclic nucleus incorporating a y-lactone ring such as in (-)- α -narcotine or (-)- β -hydrastine.² All of the synthetic routes to the phthalideisoquinolines available at the inception of the present work required the use of the somewhat inaccessible meconine or one of its analogs.^{2,3} It seemed to us that a practical route to this series could proceed from the readily available hydroxymethylpapaverine (2), itself derived from the plentiful papaverine (1).4 It is well established that the benzylic α position in papaverine is activated and lends itself to facile oxidation, and by extension it was now determined that controlled chromium trioxide in HOAc-H₂SO₄ oxidation of 2 led in 75% yield to the aromatic phthalideisoquinoline 3. Catalytic reduction of 3 with Adams catalyst in ethanol containing perchloric acid afforded in near quantitative yield and in almost equal proportions a mixture of the diastereoisomeric norphthalideisoquinolines 7 and 9 which could be separated chromatographically. Subsequent N-methylation of each of the two racemates by the formaldehyde-borohydride method furnished the required phthalideisoquinolines 8 and 10, respectively. Alternatively, the mixture of the diastereoisomeric norphthalideisoquinolines could be N-

†This conclusion requires an exchange in the assignments of the chemical shifts for H-5 and H-8 in corlumine and bicuculline made in Ref. 5. A similar change is also necessitated for cordrastine I in Ref. 3.

‡The fact that H-5 always appears near $\delta 6.6$ allows assignments for the chemical shifts of this proton in bicuculline, cordrastine II, capnoidine and cordrastine I which now become $\delta 6.64$, 6.65, 6.64 and 6.66, respectively. Bicuculline and cordrastine II belong to the erythro series and must be represented by a conformation similar to 8B. Capnoidine and cordrastine I are threo bases which exist in conformation 10B. The conformation of the alkaloid adlumine, belonging to the threo series, is also as in 10B.

methylated first, and then separated chromatographically into 8 and 10. The norphthalideisoquinolines 7 and 9 were further characterized by means of their crystalline N-acetyl amides.

The relative stereochemistry and some aspects of the conformation of the phthalideisoquinolines were initially considered by Safe and Moir who correctly established (+)-adlumine (4) to be threo and (-)- β -hydrastine (5) and (-)- α -narcotine (6) to be erythro on the basis of conclusions derived from NMR data. One of their significant observations was that the low coupling constant for H-1 and H-9 ($J_{1,9} = 3.4 - 4.3$ Hz) indicates a dihedral angle of about 50° between these two hydrogens. Another was that the chemical shift of H-2' can be used to differentiate between the threo and erythro series.

The fact that pairs of diastereoisomeric norphthalideisoquinolines as well as phthalideisoquinolines were now available to us has allowed for a firm establishment of conformations and explicit assignments of chemical shifts. A convenient starting point at this stage was the observation that, in the tetrahydrobenzylisoquinoline series, ring C lies close to the N atom if that N is secondary, but that following N-methylation ring C is found in the proximity of ring A and away from the relatively bulky N-Me group. Presently, this same steric factor, i.e. N-methylation, has also been found to prevail in the phthalideisoquinolines, so that N-methylation forces rings C and D away from the N and into proximity with ring A.

Of the four aromatic protons present in each of species 7-10, the hydrogen most distant from the γ -lactone ring, and whose chemical shift would be least subject to stereochemical or conformational considerations is H-5, and it can be seen (Table 1) that the chemical shift for H-5 remains nearly constant around $\delta 6 \cdot 6 \cdot \uparrow \pm 1$ Furthermore, the

Table 1. NMR chemical shifts of phthalideisoquinolines (δ,CDCl₃)‡

Compound	о-сн ₃			N-H N-C	н-сн3	H-5	н-8	н-1	H-9	н-2°	H-5'	
Nor-erythro 7	3.62,	3.87,	3.87,	3.90	2.06	-	6.65	6.76	4.74	5.69	5.84	7.25
Erythro &	3.75,	3.78,	3.87,	3.92	-	2.60	6.62	6.20	4.08	5.55	6.47	7.25
Nor-threo 9	3.78,	3.82,	3.89,	3.92	1.90	-	6.67	6.52	4.58	5.61	7.01	7.18
Three 10	3.72	3.77,	3.82,	3.90	~	2.69	6.65	6.32	4.09	5.62	7.00	7.17

* $J_{1,9} \approx 3.5$ Hz. H-5 and H-8 are slightly split ($^{5}1$ Hz) by interaction with H-4 and H-1, respectively, H-9 is further split ($^{5}1$ Hz) by H-2', and vice versa.

most downfield aromatic signal may be assigned in each case to H-5' which falls within the deshielding zone of the lactone carbonyl. The chemical shift for H-1 is affected by methylation on nitrogen, in both the *erythro* and *threo* series, with a resulting upfield shift of ~ 0.6 ppm (Table 1).

As observed by Safe and Moir, the signals for H-1 and H-9 may be readily recognized because of their low coupling constant, $J_{1,9} = 3.5 \,\text{Hz}$ (Table 1), so that the dihedral angle ϕ is close to 50° . In each of the four compounds, 7-10, considered here, there are only two staggered conformations (A and B) which fit this requirement.

If one considers the nor-erythro 7 and erythro 8 series first, it can be seen that the H-8 signal in erythro 8 appears upfield at $\delta 6.20$ due to shielding by ring D, so that conformation 8B must prevail. In the nor-erythro base 7, it is the H-2' and 3'-MeO signals, at $\delta 5.84$ and 3.62, respectively, that are upfield (Table 1), due to ring A shielding, and conformation 7A predominates.

The chemical shift of H-8 can also be used as a probe in establishing conformation in the nor-threo 9 and threo 10 series. This signal is relatively upfield, at $\delta 6.32$, in the case of threo 10 because of shielding by the lactone carbonyl, thus leading to the assignment of conformation 10B. It is further downfield at $\delta 6.52$ in the nor-threo base 9; no shielding is involved in this instance, and conformation 9A is paramount.

An interesting case is that of the *erythro* alkaloid narcotine (6) which incorporates a methoxyl at C-8, thus forcing the molecule into a 7A type conformation. Two opposing steric factors, namely N-methylation and substitution at C-8, can therefore influence the conformation of the phthalideisoquinoline bases. N-Methylation tends to move rings C and D towards ring A, while substitution at C-8 forces rings C and D towards the nitrogen atom, even when this nitrogen is methylated.

The realization that we had on hand the two diastereisomeric norphthalideisoquinolines 7 and 9 meant that a new route to the 13-hydroxyprotoberberines was now possible. Earlier syntheses of 13-hydroxyprotoberberines had involved either hydroboration of a 7,8-dihydroprotoberberine or LAH reduction of a phthalideisoquinoline followed by recyclization and N-demethylation.

When erythro-norphthalideisoquinoline 7 was refluxed for 5 hr with methanolic KOH, a 90% yield of the corresponding lactam alcohol 11 was obtained. Alternatively, when the threo-norphthalideisoquinoline 9 was treated with the same base under milder conditions, i.e. for 24 hr at room temperature, again a high yield of the diastereoisomeric lactam alcohol 12 was generated. If tougher conditions were used in the lactamization of 9, namely refluxing in methanolic KOH for 20 hr, a high yield of the corresponding oxyprotoberberine 15 was

obtained, resulting from facile dehydration of lactam alcohol 12 where the C-13 hydroxyl is *trans* to the C-14 hydrogen. No such dehydration was observed from the lactamization of 7.

LAH reduction of 11 and 12 yielded the required 13-hydroxyprotoberberine bases 13 and 14 respectively. Consonant with its relative stereochemistry, the 13-hydroxyprotoberberine 13 showed an NMR spectrum with H-13 at $\delta 4.76$, $J_{13,14} = 9$ Hz. This large coupling constant is in accordance with a *trans* relationship between H-13 and H-14.¹⁰

In conclusion, it can be stated that the three factors that affect the conformation of phthalideisoquinolines are (a) the relative stereochemistry at C-1 and C-9, (b) the substitution on nitrogen and (c) the substitution at C-8. The present sequence also affords a relatively short route to lactam alcohols of type 11 and 12, as well as to 13-hydroxylated tetrahydroprotoberberines.

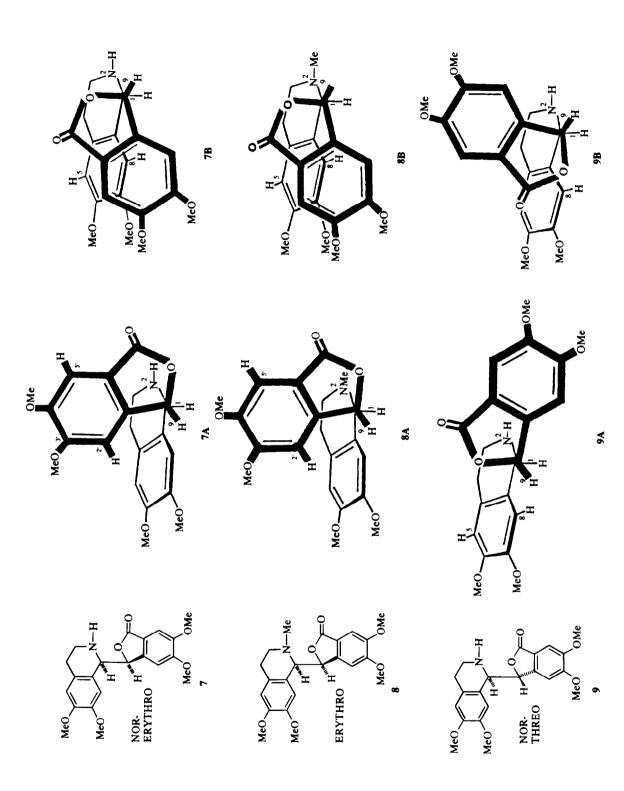
EXPERIMENTAL

Standard experimental procedures. Microanalyses were performed by Midwest Microlab, Inc., Indianapolis. M.ps are uncorrected. The NMR data is at 60 MHz in CDCl₃; and TMS was the internal standard. Mass spectra were obtained on an AEI MS-902 spectrometer. All TLC was on Merck Silica Gel-254 plates.

2'-Hydroxymethylpapaverine (2). Paraformaldehyde (6 g) was dissolved with slight heating in a mixture containing 600 ml AcOH and 40 ml conc HCl. Papaverine (20 g, 0.06 mol) was added and the mixture heated 15 hr at 60°. A large volume (600 ml) water was added, and the soln neutralized with NaHCO₃, and extracted with CHCl₃. Work-up gave 17·2 g of colorless crystals, m.p. 172-174° (benzene).*

Aromatic phthalideisoquinoline 3. Compound 2 (5 g, 0.013 mol) was dissolved in 80 ml of a soln of AcOH- H_2 SO₄- H_2 O (5:1:2), and a soln of CrO₃ (6 g, 0.06 mol) in 40 ml of AcOH- H_2 O (1:1) was added dropwise with stirring and cooling (ice-water). The dark brown mixture was stirred and cooled for an additional 20 min until the soln became dark green. The mixture was diluted with 400 ml H_2 O, and 20 g of NaHSO₃ was added portionwise with slight heating (60°). After cooling, the mixture was extracted with chloroform. The organic extract was washed with a soln of sat NaHCO₃, and then H_2 O. Work up gave a dark yellow solid which was recrystallized from EtOH, 3-8 g colorless crystals (76%), m.p. 190-192°; $\nu_{max}^{CHCl_3}$ 1760 cm⁻¹; λ_{mos}^{ErOH} 233 sh, 248, 300 and 335 nm (log ϵ 3-79, 3-10 and 2-84). High resolution mass measurement, M^+ , Found: m/e 381-1227. Calcd. for $C_{21}H_{19}$ NO₆: 381-1211.

Catalytic reduction of aromatic phthalideisoquinoline 3. A soln of 3 (200 mg, 0.52 mmol) in 50 ml EtOH containing 20 drops of 70% HClO₄ was hydrogenated over PtO₂ (50 mg) at 30 psi for 3 hr. The catalyst was removed and the solvent evaporated in vacuo; water was added and the soln neutralized with dil NHLOH. Extraction with CHCl₃ and work-up gave a mixture of the diastereoisomeric racemic 7 and 9. Preparative TLC using benzene-MeOH (4:1) afforded the nor-erythro isomer 7 (65 mg, R_f 0.41) and the nor-threo isomer 9 (60 mg, R_f 0.60).



Recrystallization of 7 from EtOH gave colorless crystals, m.p. $183-185^{\circ}$; $\nu_{\max}^{\text{CHCI}_3}$ 1750 cm^{-1} ; $\lambda_{\max}^{\text{EtOH}}$ 212, 227, 262, 295 and 310 sh nm (log ϵ 3·88, 3·82, 3·27, 3·19 and 2·98). (Found: C, 65·27; H, 6·12. Calcd. for $C_{21}H_{23}NO_6$: C, 65·44; H, 6·02%).

Recrystallization of 9 from EtOH yielded colorless crystals, m.p. 205–207°; $\nu_{\text{max}}^{\text{CNC1}}$, 1750 cm⁻¹; $\lambda_{\text{mod}}^{\text{EtOH}}$ 215, 228, 263, 295 and 308 sh nm (log ϵ 4-03, 4-08, 3-58, 3-57 and 3-38). (Found: C, 64-70; H, 6-10. Calcd. for C₂₁H₂₃NO₆-1/2 C₂H₃OH: C, 64-70; H, 6-37%).

N-Methylation of diastereoisomeric mixture of norphthalideiso-quinolines 7 and 9. The diastereoisomers 7 and 9 (170 mg, 0.44 mmol) obtained from the catalytic reduction of 3 were dissolved in 2 ml of 37% aq. formaldehyde, and the soln heated for 2 hr at 110°. Evaporation of the solvent in vacuo left a solid which was dissolved in hot MeOH. The soln was cooled in an ice bath, and excess NaBH₄ (35 mg, 0.9 mmol) was added cautiously. Stirring was continued for an additional 15 min at 0°. The mixture was then diluted with water and extracted with CHCl₃. After drying and evaporation of the solvent, the residue was separated by TLC using benzene-MeOH (4:1). The two components isolated were erythro-8 (62 mg, R_I 0.54), and threo-10 (58 mg, R_I 0.67).

Erythro 8, m.p. 157–159° (EtOH); $\nu_{\text{max}}^{\text{CHCl}_3}$ 1750 and 2780 cm⁻¹; $\lambda_{\text{max}}^{\text{EtOH}}$ 218 sh, 230, 262, 295 and 307 sh nm (log ϵ 4·20, 4·25, 3·95, 3·89 and 3·73). (Found: C, 65·90; H, 6·20, Calcd. for $C_{22}H_{25}NO_6$: C, 66·15; H, 6·31%).

Threo 10, m.p. $115-117^{\circ}$ (EtOH), $\nu_{\text{max}}^{\text{CHCl}_3}$ 1750 and 2780 cm⁻¹; $\lambda_{\text{max}}^{\text{EtOH}}$ 230, 262, 296 and 307 sh nm (log ϵ 4·13, 3·97, 3·89 and 3·78). (Found: C, 65·89; H, 6·80. Calcd. for $C_{22}H_{25}NO_6\cdot1/2C_2H_3OH$: C, 65·54, H, 6·63%).

N-Methylation of erythro-norphthalideisoquinoline 7 and of threo-norphthalideisoquinoline 9. Nor-Erythro 7 (20 mg, 0.05 mmol) was N-methylated using aq. formaldehyde (1 ml), MeOH (10 ml) and NaBH₄ (8 mg, 0.2 mmol) as above to give 18.2 mg of 8. Similarly 20 mg of 9 yielded 19.2 mg of 10.

(±)-erthro-6, 7, 3', 4'-Tetramethoxy-N-acetyl-1, 2, 3, 4-tetrahydronorphthalideisoquinoline. A soln of nor-erythro 7 (20 mg, 0·05 mmol) in one ml Ac₂O containing 8 drops of pyridine was stored for 20 hr at room temp. The solvent was evaporated, and water and CHCl₃ added. Work-up of the organic layer gave 18 mg of erythro-6,7,3',4'-tetramethoxy-N-acetyl-1,2,3,4-tetrahydronorphthalideisoquinoline, m.p. 204-205° (EtOH); $\nu_{max}^{CHCl_3}$ 1750 and 1625 cm⁻¹; λ_{max}^{EtOH} 215, 230, 265, 295 and 305 sh nm (log ϵ 4-46, 4-43, 3-90, 3-86 and 3-73); NMR δ 2-27 (3H, s, CH₃CO). (Found: C, 64-28; H, 5-98. Calcd. for C₂₃H₂₅NO₅: C, 64-62; H, 5-90%).

(±)-threo-6, 7, 3', 4'-Tetramethoxy-N-acetyl-1, 2, 3, 4-tetrahydronorphthalideisoquinoline. The above procedure was followed. From 20 mg of 9, 17 mg of the corresponding acetamide was obtained, m.p. 230–231° (EtOH); $\nu_{\text{max}}^{\text{CHCl}_3}$ 1750 and 1635 cm ¹; $\lambda_{\text{min}}^{\text{EKOH}}$ 215, 228, 265, 295 and 305 sh nm (log ϵ 4·53, 4·48, 3·91 and 3·86); NMR δ 1·90 (3H, s, CH₃CO). (Found: C, 64·25; H, 6·18. Calcd. for $C_{23}H_{25}NO_7$ ·1/2C₂H₃OH: C, 64·00; H, 6·22%).

(±)-trans-2,3,10,11-Tetramethoxy-8-oxo-13-hydroxyberbane (11). Erythro-7 (70 mg, 0.18 mmol) was dissolved in 10 ml of MeOH, and 15 mg of KOH added. The soln was refluxed for 5 hr. AcOH was added, and the soln evaporated. The residue was taken up in CHCl₃. Work-up gave 63 mg (90%) of a solid which was

recrystallized from EtOH, m.p. $210-212^\circ$; $\nu_{max}^{\text{EHCI}_3}$ 1630 cm^{-1} ; $\lambda_{max}^{\text{ENOH}_4}$ 213, 230, 270, 295 and 307 sh nm (log ϵ 4-28, 4-38, 3-81, 3-43 and 3-22). (Found: C, 65-62; H, 6-21. Calcd. for $C_{21}H_{23}NO_6$: C, 65-44; H, 6-02%).

(±)-cis-2, 3, 10, 11-Tetramethoxy-8-oxo-13-hydroxyberbane (12). Threo-9 (50 mg, 0·13 mmol) was dissolved in 10 ml of MeOH, and 11 mg of KOH added. The soln was left at room temp for 24 hr. Work-up gave 44 mg (89%) of colorless crystals, m.p. 209-211° (EtOH): $\nu_{\rm max}^{\rm CHC1}$, 1645 cm⁻¹; $\lambda_{\rm max}^{\rm ENOH}$ 212, 230, 275, 297 and 306 sh nm (log ϵ 3·93, 3·96, 3·34, 3·30 and 3·11). (Found: C, 65·24; H, 6·00. Calcd. for C₂₁H₂₅NO₆: C, 65·44; H, 6·02%).

(±)-trans-2, 3, 10, 11-Tetramethoxy-13-hydroxyberbane (13). Lactam 11 (20 mg, 0.05 mmot) in 3 mt THF was refluxed with LAH for 8 hr to yield 15 mg (78%) crystals, m.p. 197–199° (MeOH); $\lambda_{\text{max}}^{\text{EIOH}}$ 212, 230 and 290 nm (log ϵ 4·17, 3·93 and 3·48). (Found: C, 66·60; H, 6·68. Calcd. for C₂₁H₂₅NO₅·1/2CH₃OH: C, 66·66; H, 6·97%).

(±)-cis-2, 3, 10, 11-Tetramethoxy-13-hydroxyberbane (14). Lactam 12 was reduced as above in 81% yield to give crystals, m.p. 198–200° (MeOH-ether); λ_{max}^{EIOH} 213, 233 sh and 288 nm (log ε 4·07, 3·56 and 3·17). (Found: C, 67·68; H, 6·95. Calcd for C₂₁H₂₅NO₅: C, 67·90: H, 6·78%).

Oxyprotoberberine 15. (\pm)-cis-12 (20 mg, 0.5 mmol) was dissolved in 10 ml of MeOH containing 4 mg of KOH, and the soln refluxed 20 hr under N₂. AcOH was added, the solvent evaporated, and the residue extracted with benzene. Work-up gave nearly colorless prisms, 15 mg (78%), m.p. 195-197° (CHCl₃-ether); $\nu_{\text{max}}^{\text{CHCl}_3}$ 1635 cm⁻¹; $\lambda_{\text{max}}^{\text{ROM}}$ 232, 265, 338 and 350 sh nm (log e 3-90, 3-73, 3-59 and 3-46); NMR 82-92 (2H, t, J = 6 Hz, CH₂), 3-93 (3H, s, OCH₃), 3-97 (3H, s, OCH₃), 4-00 (6H, s, 2OCH₃), 4-37 (2H, t, J = 6 Hz, CH₂N), five arom. H s at 6-72, 6-82, 6-92, 7-25 and 7-80. High resolution mass measurement, M*: Found: m/e 367-1397.

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Calcd. for $C_{21}H_{21}NO_5$: 367-1420.

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